12/C GnD 10/4/02

IN THE UNITED STATES PATENT AND TRADEMARK OFFICE

APPLICANT: Kline & Sanders	§	**ART UNIT: 1644**
	§	
FILED: February 10, 2000	§	**EXAMINER:**
	§	**Nolan, P.**
SERIAL NO.: 09/501,912	§	
	§	**DOCKET:**
FOR: Targeted Destruction of Pests	§	**D6017CIP**

The Assistant Commissioner of Patents and Trademarks
BOX NON-FEE AMENDMENT
Washington, DC 20231

RESPONSE UNDER 37 C.F.R. § 1.111

Dear Sir:

In response to the Office Action mailed April 22, 2002, please enter the following amendments and remarks. Reconsideration of the pending claims is respectfully requested.

AMENDMENTS

IN THE SPECIFICATION:

Please replace the paragraph beginning on page 25, line 11, with the following rewritten paragraph:

cDNA synthesis RNA was isolated from mouse spleens (1/2 spleen from mice immunized with midgut preparations from imported fire ant queens as described in Example 1) using the

C1

48

C

guanidium isothiocyanate method. cDNA was prepared from 5 micrograms of RNA with oligo $(dT)_{16}$ as a primer. Reverse transcriptase, nucleotides, and buffers were purchased from PERKIN ELMER (RNA PCR Kit, Branchburg, New Jersey) and were used according to the instructions provided by the manufacturer. Fd and L chain cDNA were amplified by PCR. The 5' primers used were Light chain (GTGCCAGATGTGAGCTCGTGATGACCCAGTCTCCA, SEQ ID NO:1), V heavy chain a (AGGTCCAGCTGCTCGAGTCTGG, SEQ ID NO:2), VHb (AGGTCCAGCTGCTCGAGTCAGG, SEQ ID NO:3), V heavy chain c (AGGTCCAGCTTCTCGAGTCTGG, SEQ ID NO:4), and V heavy chain D (AGGTCCAGCTTCTCGAGTCAGG, SEQ ID NO:5) which introduced restriction sites (Sac I for light chains and XHO 1 for heavy chains) that facilitate their directional cloning into pComb 3. The 3' primers used were k chain (TCCTTCTAGATTACTAACACTCTCCCCTGTTGAA, SEQ ID NO:6), C heavy 1 (AGGCTTACTAGTACAATCCCTGGGCACAAT, SEQ ID NO:7), thereby the k chain primer introduced an Xba 1 site and the heavy chain primer introduced a Spe 1 site. General conditions for PCR were Taq polymerase (Perkin Elmer, Branchburg, New Jersey) at 2.5 U/100-microliter reaction mixtures, 200 micromolar deoxynucleoside triphosphates, 1 millimolar $MgCl_2$, 5 microliters of cDNA per 100 microliters of reaction mixture, 150 ng of 5' primer and